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Dairy Thermodurics

Module 4: Persistence of Thermodurics in Dairy Processing Plants

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Content

- ▶ Are thermodurics in raw milk a real cause of worry for the milk processors?
- ▶ What makes thermodurics so persistent?
- ▶ The concept of bacterial biofilms



Why thermodurics are so difficult to eliminate from the processing environment?

- ▶ Resistant to thermal processing such as industrial pasteurization
- ▶ Many form spores having greater resistance to heat and chemicals
- ▶ Wide temperature growth range
- ▶ Thermoduric thermophiles exhibit a faster growth rate
 - ▶ generation time 15-20 min
- ▶ Readily form biofilms

(Flint et al, 1999 & 2001; Parker et al, 2001, Scott et al, 2007; Burgess et al, 2010; Hassan et al., 2010)



Persistence; The biofilm angle

- ▶ Biofilms are congregation of sessile bacteria attached to a surface
- ▶ Bound together with extracellular polymeric substance (EPS)
 - ▶ Proteins, lipids, and polysaccharides
- ▶ Occur in areas of initial mineral deposits or organic matter buildup
 - ▶ Soiling within milking or processing equipment
- ▶ Biofilms formed by thermotolerant bacteria
 - ▶ Both spores and vegetative cells can attach to stainless steel and fouled surfaces
 - ▶ Foulant or biofilms may protect spores and vegetative cells against CIP chemicals

(Costerton et al., 1994; Hassan et al., 2010, Anand et al., 2012)

Areas prone to thermoduric biofilms

- ▶ Following are the major areas prone to biofilms in the processing plants
 - ▶ Pasteurizer heat exchange plates; regeneration section
 - ▶ Cream separators
 - ▶ Regeneration section of evaporators
 - ▶ Preheater of evaporation unit in spray drying
 - ▶ Membrane applications in milk and whey processing
 - ▶ Cream heater and recycle loops in butter plants
- ▶ Remain ~45 - 60°C/ 113 - 140°F
 - ▶ Support growth of thermodurics, especially thermophiles
- ▶ Other problem areas
 - ▶ Rubber seals, corners, hard to clean areas

(Scott, 2007; Burgess, et al., 2009)



Factors influencing biofilm formation

- ▶ The ratio of planktonic (free-floating) cells to biofilm embedded cells is a function of several interrelated factors
 - ▶ Types of bacteria
 - ▶ Surface energetics
 - ▶ Materials of construction
 - ▶ Microtopography of surfaces
 - ▶ Hydraulic factors
 - ▶ Biofilm chemistry



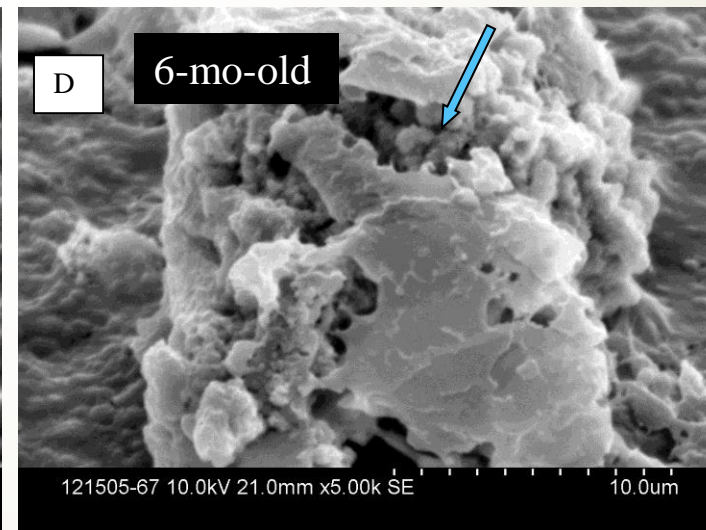
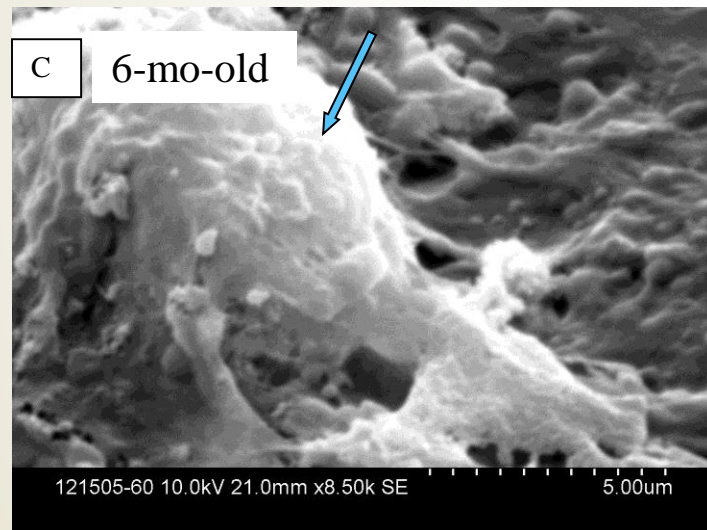
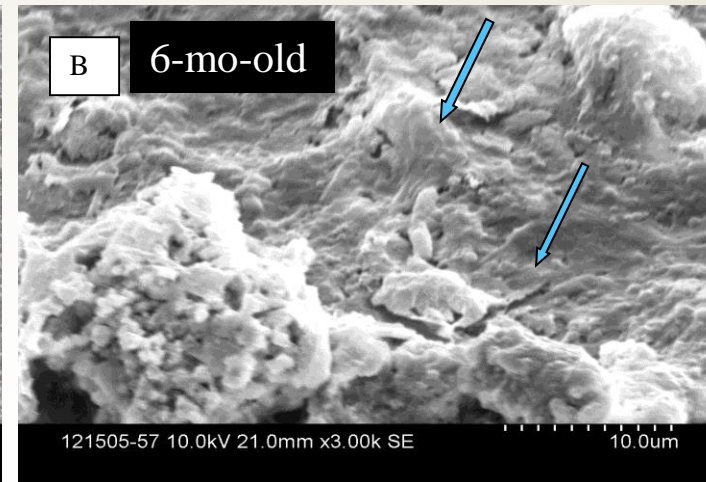
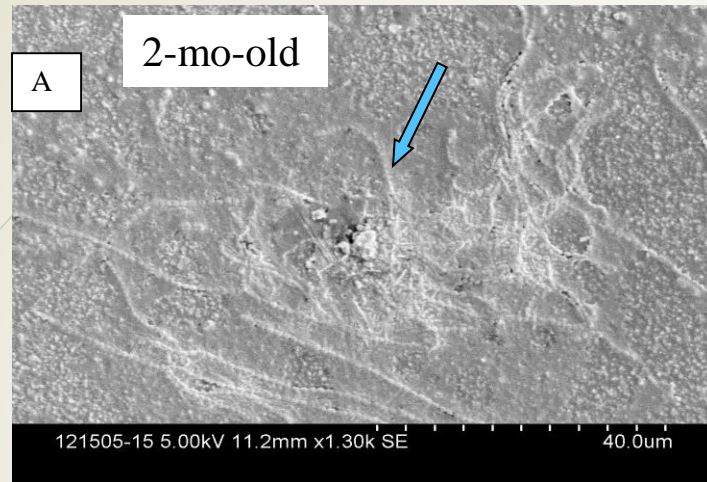
Stages of a typical biofilm formation

- ▶ Stage I: (within 5-10 sec) surfaces are coated with milk proteinaceous components and calcium phosphate
- ▶ Stage II: (within 6-8 hours) primary colonization of bacterial cells with the substratum
 - ▶ Extrapolysaccharides (EPS) appears to support this process
- ▶ Stage III: (within 10-12 hours) irreversible biofilms
 - ▶ Formation of mature biofilms with embedded bacteria in the organic film matrix



SEM showing biofilm structures on used RO membranes

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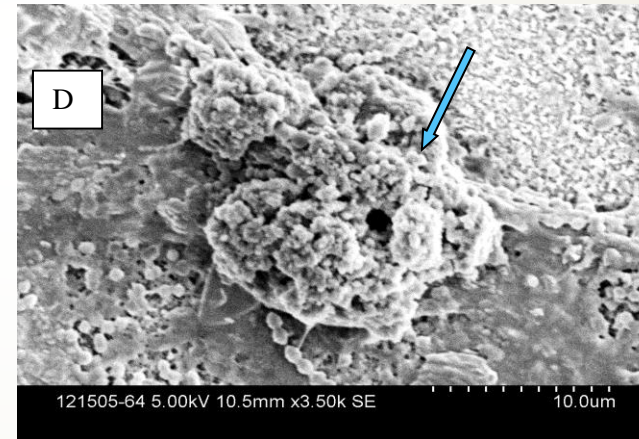
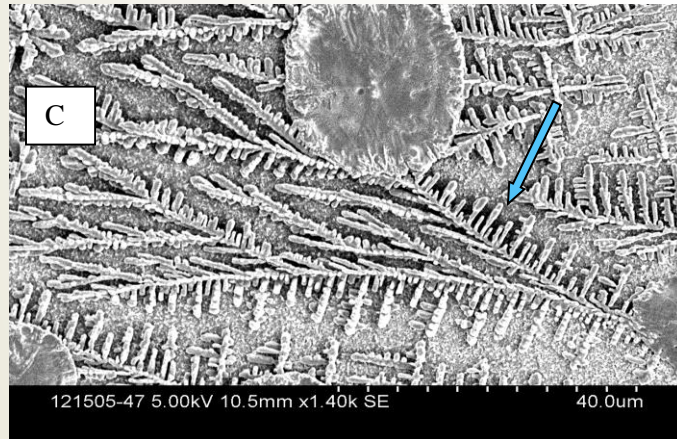
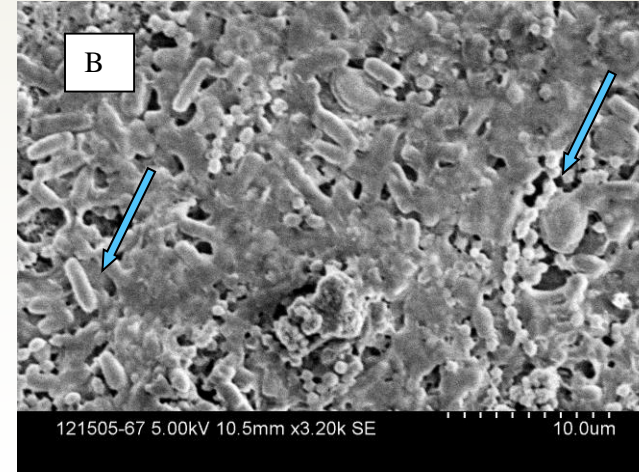


2mo-old membrane showing long chains of cocci (A), 6mo-old membrane showing hill and valley structure (B), Mushroom like structure (C), and Ruptured mushroom (D). (*Hassan et al., 2010*)

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SEM showing different biofilm structures on aged membranes

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12mo-old membrane showing monolayer of rods and cocci connected by EPS (A), 14mo-old membrane showing rods and cocci embedded in biofilm matrix (B), Distinct biofilm structures (C&D) (*Hassan et al., 2010*)

Evidence of cross contamination of whey retentate

- ▶ In a study the presence of the same *Bacillus* sp. was reported in both membrane biofilm and the corresponding whey retentate
- ▶ The absence of this species in the feed whey confirmed the cross contamination of the retentate from the biofilm embedded bacteria

(Anand et al., 2012)

Resistance of biofilm embedded bacteria to regular CIP

- ▶ Biofilms embedded bacteria are more resistant to cleaning as compared to their planktonic counterparts
- ▶ The existing CIP protocol from a whey concentration plant was tested against the biofilm consortia isolates obtained from used whey RO membranes (2 to 14 months old)
- ▶ The six steps CIP process was based on; alkali, surfactant, acid, enzyme, a second surfactant, and a weekly sanitizer application
- ▶ The individual isolates showed a great variability in their sensitivity towards different chemical treatments
 - ▶ In general, the spore forming *Bacillus* spp. showed greater resistance as compared to the other genera such as *Enterococcus*, *Escherichia*, *Streptococcus*, *Staphylococcus*, and *Klebsiella*

(Anand and Singh, 2013)



Making CIP more effective using certain enzymes as biocleaners

- ▶ Different enzymes including a protease, lipase, lactase, α -glucosidase, and β -galactosidase were selected to evaluate their ability to degrade the biofilm matrix
 - ▶ A 24 hour old membrane biofilm formed by a resistant *Bacillus* isolate was used for this evaluation
 - ▶ β -galactosidase showed highest reduction in biofilm counts

(Anand and Singh, 2013)

Credits

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